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Editorial



It's time to make melatonin a useful tool in improving cardiac rhythmicity

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Melatonin has a profound effect on the pacemaker of the central mechanism of a heart. 1-3 That this heart is in a fly should not affect the importance of this fact. Drosophila melanogaster has done yeoman service across biology beginning with providing the foundation of modern genetics.^{4,5} Multiple Nobel prizes, the most recent in 2017, have gone to those working with the fly.5 This fly work includes cardiology. The fly heart has been studied extensively. 6-10 Its physiology has been described in detail and this includes an understanding of basic hormonal control. 11,12 The sarcolemmal pacemaker's central ion channels are well established, and the physiology and components of the companion cytosolic pacemaker have also been studied in detail. 11-18 The utility of the fly genome in the study of the mammalian heart has been clearly proven. Long QT syndrome, a congenital problem in which individuals suffer a delay in cardiac repolarization, 19-21 has been found to be a result of a mutation in a gene originally found in the fly.22-25 The human Ether a go Related Gene, or hERG is a homolog of the fly gene seizure and cDNA from this gene was used to locate its human relative.²² There have been over 1600 human disease genes uncovered to date and nearly three quarters of these have Drosophila homologs. About 500 are highly conserved and known to be functionally equivalent in both organisms.²⁶ With this mind, cardiologists might well look to the fly for answers to basic questions about underlying heart physiology, genetics, and pathology.

Melatonin makes the fly heartbeat extremely regular. 1-3 It cannot be emphasized too strongly that the alteration in heartbeat we observe is unprecedented. We showed this is not an artifact of an increase in rate.1 Melatonin is commonly used in humans to prevent damage by reactive oxygen species (ROS) during reperfusion after myocardial infarction.²⁷ Reports of increased heartbeat regularity attribute this to the antioxidant effects.^{27–31} However, this was shown not to be the case in the fly; ascorbic acid has no such effect in Drosophila.1 It is important to note here that the profound change in the signal to noise ratio in the fly work is against a background of a fairly irregular heartbeat in the normal fly. 1-3 Mammalian hearts are also not particularly regular under normal conditions, to the point that there has been considerable work done to find out if this is a result of the underlying oscillator being fundamentally chaotic. 32-34 In humans, an abnormally regular heartbeat can be a sign of congestive heart failure.32-34

One crucial observation from these studies is that melatonin can yield normal wild-type heart function in flies carrying a mutation in one of the genes encoding a central ion channel in the sarcolemal pacemaker, *slowpoke*, which normally shows a weak, highly erratic heartbeat.¹⁴ The beating observed after melatonin application is equal to the extremely abnormal high regularity seen in wild-type. This is, by itself, a provocative finding. It would be comparable to a car being made to run normally with an engine which has broken a

camshaft. A possible explanation is at hand. Considerable evidence has accumulated that there is a second cardiac pacemaker in the cytosol termed the LCR, for Local Calcium Release.^{35–39} Evidence for such an oscillator is strong. One compelling observation is that pacemaking can continue in a vertebrate heart cell when the plasma membrane of the cell is voltage clamped!³⁵ This oscillator depends on Calcium currents passing through the membrane of the sarcoplasmic reticulum.^{35–39} There are two central ion channels: one is the ryanodine receptor efflux other allowing and the Sarcoplasmic/Endoplasmic Calcium ATPase (SERCA) effecting reuptake. 40-42 Mutations in the genes encoding these channels affect heart function considerably 40-42 and melatonin is relatively ineffective in ameliorating the effects.³ An immediate hypothesis is that melatonin is fundamentally affecting the communication between these two pacemakers. If the sarcolemmal oscillator fails, responsibility would shift entirely to the LCR system. A switch to the LCR oscillator would also explain the wildtype results. 1-3

It is of importance to learn how melatonin works. Two possibilities present themselves. The effect could be directly on the pacemaker through interactions with the ion channels comprising the oscillator. A second control pathway possibility would be through binding with a receptor. The latter is initially favored by the kinetics of the alteration. We observed that the switch between normal mode and hyper regularity occurs sharply, literally from one beat to the next after an interval post injection.¹ With this in mind, we tested melatonin receptor agonists and antagonists with positive results. Luzindole is a melatonin receptor antagonist, and it is effective in interdicting melatonin's action. In contrast, 2-[125I] iodomelatonin, a melatonin agonist, is even more effective in increasing the rhythmicity than melatonin. The next step was to identify the receptor. We used RNAi knockout techniques to probe likely orphan G Protein Coupled Receptor (GPCR) genes for a candidate. Knocking out the function of the CG4313 orphan completely eliminated the melatonin response.1 This finding makes it almost certain that the mechanism is receptor mediated.

The flies have done their job. There is ample reason to hypothesize that given the similarities between the systems, these findings could be extended to the mammalian heart. The reported examples of increased regularity in hearts given melatonin to preclude reperfusion damage after infarct are strong evidence this is the case, especially in light of our finding that the antioxidant ascorbic acid has no effect on rhythmicity. The basic work needs to be done, and translational work would not be far behind.

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Conflicts of Interest

The author declares that there are no conflicts of interest.

References

- 1. VanKirk T, Powers E, Dowse HB. Melatonin increases the regularity of cardiac rhythmicity in the *Drosophila* heart in both wild–type and strains bearing pathogenic mutations. *J Comp Physiol B*. 2017;187(1):63–78.
- Dowse H, VanKirk T. Drosophila as a Model System for Cardiology: The Case of Melatonin and Heartbeat Regularity. *Medical Research Archives*. 2022;10(5).
- 3. Dowse H, T VanKirk. Communication Between the Plasma Membrane and Cytosolic Cardiac Pacemakers: A Role for Melatonin? 2023;11(2). *Medical Research Archives*.
- Roberts D. Drosophila melanogaster: the model organism. Entomologia Experimentalis et Applicata. 2006;121:93–103.
- Yamaguchi M, Yoshida H. Drosophila as a Model Organism. Drosophila models for human diseases In: Advances in Experimental Medicine and Biology. Springer. 2018;1–10.
- 6. Rizki TM. The circulatory system and associated cells and tissues. In: Ashburner M; WT, editor. *The Genetics and Biology of Drosophila*. London: Academic Press. 1978;1839–1845.
- Bodmer R, Venkatesh T V. Heart development in Drosophila and vertebrates: Conservation of molecular mechanisms. Dev Genet. 1998;186:181– 186
- 8. Curtis NJ, Ringo JM, Dowse HB. Morphology of the pupal heart, adult heart, and associated tissues in the fruit fly, Drosophila melanogaster. *J Morphol*. 1999;240:225–235.
- 9. Bier E, Bodmer, R. Drosophila, An emerging model for cardiac disease. *Gene*. 2004;342:1–11.
- Taghli–Lamallem O, Plantie E, Jagla K. Drosophila in the heart of understanding cardiac diseases: Modeling channelopathies and cardiomyopathies in the fruitfly. J Cardiovasc Dev and Dis. 2016;3:7–28.
- Johnson E, Ringo J, Dowse H. Modulation of *Drosophila* heartbeat by neurotransmitters. *J Comp Physiol B*. 1997;167:89–97.
- 12. Johnson E, Ringo J, Dowse H. Native and heterologous neuropeptides are cardioactive in Drosophila melanogaster. *J Insect Physiol.* 2000;46:1229–1236.

- Dowse H, Ringo J, Power et al. A congenital heart defect in Drosophila caused by an action–potential mutation. *J Neurogenet*. 1995;10:153–168.
- 14. Johnson E, Ringo J, Dowse H, et al. Genetic and pharmacological identification of ion channels central to the *Drosophila* cardiac pacemaker. *J. Neurogenet*. 1998;12:1–24.
- Sullivan K, Scott K. The ryanodine receptor is essential for larval development in *Drosophila* melanogaster. Proc Natl Acad Sci. 2000;97:5492– 5497.
- Sanyal S, Consoulas H, Kuromi H, et al. Analysis of conditional paralytic mutants in Drosophila sarco– endoplasmic reticulum Calcium ATPase reveals novel mechanisms for regulation of membrane excitability. *Genetics*. 2005;169:737–750.
- 17. Sanyal S, Jennings T, Dowse H. et al. Conditional mutations in SERCA, the sarco–endoplalsmic reticulum Ca²⁺–ATPase, alter heart rate and rhythmicity in Drosophila. *J Comp Physiol B*. 2005;176:253–263.
- Abraham M, Wolf M. Disruption of sarcoendoplasmic reticulum calcium ATPase function in *Drosophila* leads to cardiac dysfunction. *PLOS ONE*. 2013;8:e77785.
- Curran M, Splawski I, Timothy K, et al. A molecular basis for cardiac arrhythmia: HERG mutations cause long QT syndrome. Cell. 2006;1995;80:795–803.
- Sanguinetti M, Jiang C, Curran M, et al. A mechanistic link between an inherited and an ac– quired cardiac arrythmia: HERG encodes the I_{kr} potassium channel. Cell. 1995;81:299–307.
- 21. Sanguinetti M, Curran M. Spectrum of HERG K⁺-channel dysfunction in an inherited cardiac arrhythmia. *Proc Natl Acad Sci USA*. 1996;93:2208–2212.
- 22. Warmke J, Ganetzky B. A family of potassium channel genes related to *eag* in *Drosophila* and in mammals. *Proc Natl Acad Sci USA*. 1994;91:3438–3442.
- 23. Jackson F, Wilson S, Strichartz G. et al. Two types of mutants affecting voltage–sensitive sodium channels in Drosophila melanogaster. *Nature*. 1994;308:189–191.
- 24. Wang X, Reynolds E, Deak P. The *seizure* locus encodes the *Drosophila* homolog of the HERG potassium channel. *J Neurosci.* 1997;17:882–890.
- 25. Warmke J, Ganetzky B. A family of potassium channel genes related to *eag* in *Drosophila* and in mammals. *Proc Natl Acad Sci USA*. 1994;91:3438–3442.
- Taghli-Lamallem O, Plantie E, Jagla K. *Drosophila* in the heart of understanding cardiac diseases: Modeling channelopathies and cardiomyopathies in the fruitfly. *J Cardivasc Dev Dis.* 2016;3(7).
- Tan DX, Reiter RJ, et al. Ischemia/perfusion induced arrhythmia in the isolated rat heart: prevention my melatonin. *J Pineal Res.* 1998;25:184–191.

- 28. Tan DX, Reiter RJ, et al. Ischemia/perfusion induced arrhythmia in the isolated rat heart: prevention my melatonin. *J Pineal Res.* 1998;25:184–191.
- Bertuglia S, Reiter RJ. Melatonin reduces ventricular arrhythmias and preserves capillary perfusion during ischemia-reperfusion events in cardiomyopathic hamsters. J Pineal Res. 2007;42:55–63.
- 30. Diez ER, Prados LV, Carrión A, et al. A novel electrophysiologic effect of melatonin on ischemia/reperfusion–induced arrhythmias in isolated rat hearts. *J. Pineal Res.* 2009;46:155–160.
- Benova T, Knezl V, Viczenczova C, et al. Acute antifibrillating and defibrillating potential of atorvastatin, melatonin, eicosapentaenoic acid and docosahexaenoic acid demonstrated in isolated heart model. *J Physiol Pharmacol*. 2015;66:83–89.
- 32. Goldberger L. Heartbeat Chaotic or Homestatic? *Physiology*. 1991;6:87–91.
- Wu G-Q, Arzeno NM, et al. Chaotic Signatures of Heart Rate Variability and Its Power Spectrum in Health, Aging and Heart Failure. *PLoS ONE*. 2009;4:e4323.
- Glass L. Introduction to Controversial Topics in Nonlinear Science: Is the Normal Heart Rate Chaotic? *Chaos.* 2009;19:028501.
- 35. Vinogradova T. Rhythmic Ryanodine Receptor Ca²⁺ releases during diastolic depolarization of sinoatrial pacemaker cells do not require membrane depolarization. *Circ Res.* 2004:94:802–809.
- Lakatta e, DiFrancesco D. JMCC Point-Counterpoint: What keeps us ticking, a funny current, a Calcium clock, or both? *J Mol Cell Cardiol*. 2009;47(2):157–170.
- 37. Maltsev V, Lakatta E. Synergism of coupled subsarcolemmal Ca²⁺ clocks and sarcolemmal voltage clocks confers robust and flexible pacemaker function in a novel pacemaker cell model. *Am J Physiol*. 2009;296:H594–H615.
- Lakatta E, Maltsev V, Vinogradova T. A coupled system of intracellular Ca²⁺ clocks and surface membrane voltage clocks controls the timekeeping mechanism of the heart's pacemaker. *Circulation Research*. 2010;106:659–673.
- 39. Wolk R. Arrhythmogenic mechanisms in left ventricular hypertrophy. *Europace*. 2000;2:216–223.
- Sullivan K, Scott K. The ryanodine receptor is essential for larval development in Drosophila melanogaster. *Proc Natl Acad Sci USA*. 2000;97:5492–5497.
- Sanyal S, Consoulas H, Kuromi H, et al. Analysis of conditional paralytic mutants in Drosophila sarco– endoplasmic reticulum Calcium ATPase reveals novel mechanisms for regulation of membrane excitability. *Genetics*. 2005;169:737–750.
- 42. Sanyal S, Jennings T, Dowse H. et al. Conditional mutations in SERCA, the sarco–endoplalsmic reticulum Ca²⁺–ATPase, alter heart rate and rhythmicity in Drosophila. *J Comp Physiol B*. 2005;176:253–263.